Journal of Veterinary Internal Medicine

J Vet Intern Med 2016;30:813-818

# A Homozygous *RAB3GAP1:c.743delC* Mutation in Rottweilers with Neuronal Vacuolation and Spinocerebellar Degeneration

T. Mhlanga-Mutangadura, G.S. Johnson, A. Ashwini, G.D. Shelton, S.A. Wennogle, G.C. Johnson, K. Kuroki, and D.P. O'Brien

**Background:** A variety of presumed hereditary, neurologic diseases have been reported in young Rottweilers. Overlapping ages of onset and clinical signs have made antemortem diagnosis difficult. One of these diseases, neuronal vacuolation and spinocerebellar degeneration (NVSD) shares clinical and histological features with polyneuropathy with ocular abnormalities and neuronal vacuolation (POANV), a recently described hereditary disease in Black Russian Terriers (BRTs). Dogs with POANV harbor mutations in *RAB3GAP1* which codes for a protein involved in membrane trafficking.

**Hypothesis:** Rottweilers with NVSD will be homozygous for the *RAB3GAP1:c.743delC* allele associated with POANV in BRTs.

Animals: Eight Rottweilers with NVSD confirmed at necropsy, 128 Rottweilers without early onset neurologic signs, and 468 randomly selected dogs from 169 other breeds.

Methods: Retrospective case-control study. Dogs were genotyped for the *RAB3GAP1:c.743delC* allele with an allelic discrimination assay.

**Results:** All 8 NVSD-affected dogs were homozygous for the *RAB3GAP1:c.743delC* allele while the 128 NVSD-free Rottweilers were either homozygous for the reference allele (n = 105) or heterozygous (n = 23) and the 468 genotyped dogs from other breeds were all homozygous for the reference allele.

**Conclusions and Clinical Importance:** The *RAB3GAP1:c.743delC* mutation is associated with a similar phenotype in Rottweilers and BRTs. Identification of the mutation permits a DNA test that can aid in the diagnosis of NVSD and identify carriers of the trait so that breeders can avoid producing affected dogs. Disruption of membrane trafficking could explain the neuronal vacuolation seen in NVSD and other spongiform encephalopathies.

Key words: Canine; Molecular genetics; Peripheral nervous system disorders; Rab GTPase; Spongiform encephalopathies; Warburg micro syndrome.

A number of presumed hereditary neurologic diseases occur in young Rottweiler dogs.<sup>1–9</sup> As discussed in reviews of these diseases,<sup>10,11</sup> there is considerable overlap in clinical signs and ages of onset which can make antemortem differentiation of the conditions difficult. The development of DNA tests for the mutations associated with specific diseases can help diagnose dogs with those diseases.<sup>12</sup> Recently a deletion in *RAB3GAP1* was identified in Black Russian Terriers (BRTs) with a juvenile onset laryngeal paralysis and polyneuropathy.

From the Department of Veterinary Pathobiology, College of Veterinary Medicine, University of Missouri, Columbia, MO (Mhlanga-Mutangadura, G.S. Johnson, Ashwini, G.C. Johnson, Kuroki); the Department of Pathology, University of California San Diego, La Jolla, CA (Shelton); the Department of Clinical Sciences, Colorado State University, Fort Collins, CO (Wennogle); and the Department of Veterinary Medicine and Surgery, College of Veterinary Medicine, University of Missouri, Columbia, MO (O'Brien).

The work was performed at the University of Missouri, University of California San Diego, and Colorado State University

Corresponding author: D.P. O'Brien, Comparative Neurology Program, College of Veterinary Medicine, University of Missouri, 900 East Campus Drive, Columbia, MO 65211; e-mail: obriend@missouri.edu.

Submitted November 24, 2015; Revised December 28, 2015; Accepted February 11, 2016.

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DOI: 10.1111/jvim.13921

## Abbreviations:

BRT	Black Russian Terrier			
BSE	bovine spongiform encephalopathy			
CJD	Creutzfeldt-Jakob disease			
GAP	GTPase activator protein			
GEF	guanine exchange factor			
NVSD	neuronal vacuolation and spinocerebellar degeneration			
POANV	polyneuropathy with ocular abnormalities and			
	neuronal vacuolation			
WARBM	Warburg micro syndrome			

Further investigation disclosed additional features of the disease which is now called polyneuropathy with ocular abnormalities and neuronal vacuolation (POANV).<sup>13</sup> A SINE insertion in *RAB3GAP1* was also reported in Alaskan Huskies with POANV.<sup>14</sup> Similarities between the phenotypes of POANV and one of the previously reported diseases in young Rottweilers, neuronal vacuolation, and spinocerebellar degeneration (NVSD),<sup>7</sup> prompted us to determine if the same mutation is found in dogs with NVSD.

## **Materials and Methods**

Medical records of 8 Rottweilers diagnosed with NVSD were reviewed. Necropsies were performed on 4 dogs. Brain and other tissues collected at the necropsy were fixed in 10% formalin and histopathology examination was performed after routine histologic processing using H&E, LFB/PAS and Bielschowsky stains. In one of these dogs, fresh and fixed biopsies collected from the quadriceps, cranial tibial, and the cricoarytenoideus dorsalis muscles as well as the recurrent laryngeal and common peroneal nerves were sent by courier service to the Comparative Neuromuscular Lab at University of California San Diego. Upon receipt, the unfixed muscles were flash frozen in isopentane precooled in liquid nitrogen and processed with a standard panel of histochemical stains and reactions.<sup>15</sup> The fixed muscle was processed into paraffin by standard procedures and the fixed nerves resin embedded as previously described.<sup>16</sup>

EDTA-anticoagulated blood samples or buccal swab samples<sup>a</sup> were collected from these dogs and from 128 other Rottweilers that had no recorded neurologic signs at 1 year of age. Four of the sampled NVSD-free Rottweilers were diagnosed with laryngeal paralysis, polyneuropathy, or both between 3 and 10 years of age. DNA was prepared from these samples by previously described methods.<sup>17,18</sup> An additional 468 DNA samples from 169 other dog breeds were randomly selected from the University of Missouri Animal DNA Depository. The DNA samples from individual dogs were genotyped at *RAB3GAP1:c.743* with a TaqMan allelic discrimination assay<sup>19</sup> as previously described.<sup>13</sup> All studies were approved by the Animal Care and Use Committee of the University of Missouri and conducted with informed consent of the owners.

## Results

## **Phenotype**

Eight Rottweilers presented for respiratory distress at 3–4 months of age and 7 were diagnosed with laryngeal paralysis. Other reported signs were pelvic limb sensory ataxia and weakness (n = 8), cerebellar ataxia (n = 2), megaesophagus (n = 2), cataracts (n = 3), microphthalmia (n = 2), and persistent pupillary membranes (n = 1). Histopathology of the brain in the 4 dogs examined revealed frequent intracytoplasmic vacuolation within neurons confirming a diagnosis of NVSD (Fig 1). Vacuoles were found in the following neu-



Fig 1. Neurons showed single or multiple large vacuoles characteristic of NVSD (H&E stain, bar =  $50 \mu m$ ).

roanatomic locations in the 4 dogs that were necropsied: cerebellar cortex (4/4), cerebellar roof nuceli (3/4), cuneate nuclei (4/4), gracilis nuclei (2/4), hypoglossal nuclei (2/4) inferior olives (2/4), vestibular nuclei (2/4), striate nuclei (1/4), substantia nigra (1/4), retina (1/4), and spinal cord (1/4). Irregular loss of cerebellar Purkinje cells with empty basket cell processes were prominent in all dogs, and occasional Purkinje cells had axonal torpedoes (Fig 2). Histopathology of peripheral nerve and muscle was performed on 1 dog (Fig 3). The overall density of myelinated fibers was subjectively appropriate in the common peroneal nerve (Fig 3A) with a decrease in the expected population of large caliber nerve fibers and an increase in the population of small caliber fibers consistent with mild axonal degeneration and presumptive regeneration. Myofiber size in the limb muscles was generally small (Fig 3C) with retention of a normal polygonal shape. In contrast, marked nerve fiber loss was evident in the recurrent laryngeal nerve (Fig 3B) with myofiber loss evident in the cricoarytenoideus dorsalis muscle (Fig 3D) consistent with more marked axonal degeneration.

## Genotype

All 8 Rottweilers with NVSD were homozygous for the *RAB3GAP1:c.743delC* variant allele; whereas the 128 NVDS-free Rottweilers were either heterozygotes (n = 23) or homozygous for the reference allele (n = 105). The 4 Rottweilers who presented with laryn-



Fig 2. The cerebellar cortex contained segments where only empty Basket cell processes are apparent. The black arrow shows a residual Purkinje cell body (bar =  $100 \mu$ m). Inset: A few Purkinje cells had swollen axons. This neuron is next to an empty basket. (Bielschowsky stain, bar =  $50 \mu$ m).



**Fig 3.** Muscle and nerve biopsies from a 3-month-old male Rottweiler puppy with a confirmed homozygous *RAB3GAP1:c.743delC* mutation. (A) Resin embedded 1- $\mu$ m section from the common peroneal nerve showing a subjectively normal overall population of nerve fibers with a decreased population of large caliber fibers and an increased population of small caliber fibers. (B) In comparison, similar sections from the recurrent laryngeal nerve show marked loss of all calibers of myelinated nerve fibers consistent with chronic axonal degeneration. (Paraphene-lenediamine stain, bar = 50  $\mu$ m for both A and B). (C) Paraffin-embedded section from the vastus lateralis muscle showing generalized myofiber atrophy or hypotrophy with retention of the normal polygonal myofiber shape. (D) For comparison, myofiber loss was evident in the cricoarytenoideus dorsalis muscle with wide separation of remaining muscle fascicles. (H&E stain, bar = 50  $\mu$ m for both C and D).

geal paralysis, polyneuropathy, or both at >1 year of age were all homozygous for the reference allele. All 468 dogs representing 169 breeds other than Rottweiler or BRT were homozygous for the reference allele.

## Discussion

The RAB3GAP1:c.743delC variant previously identified in the homozygous state in BRTs with POANV<sup>13</sup> also occurred as a homozygous variant in all 8 of the NVSD-affected Rottweilers in our study. None of the normal Rottweilers or the Rottweilers with signs beginning at >1 year of age were homozygous for the variant. As previously reported<sup>13</sup> the variant was not detected in randomly selected samples from purebred dogs of other breeds besides the BRT. It is likely that the same founder mutation event was the source of the RAB3GAP1:c.743delC allele in both Rottweilers and BRTs. According to the BRT Club of America website (http://www.brtca.org/brt-information.html accessed November 16, 2015), the BRT breed was developed by interbreeding Rottweilers with other selected breeds during the 1930s in a military kennel near Moscow. The goal was to create a new Russian breed for the national

security force that was large and rugged enough to endure the harsh Siberian winters. The mutation predicts a premature stop codon and a truncated gene product *RAB3GAP1:p.P248Lfs4\** missing 730 C-terminal amino acids, including the catalytic domain. Thus, it is doubtful that the truncated gene product retains biological activity.

*RAB3GAP1* codes for the catalytic subunit that combines with a noncatalytic subunit encoded by *RAB3-GAP2* to form Rab3GAP. Rab3GAP was first recognized as a GTPase activator protein (GAP) that greatly enhances the inherent GTPase activity of Rab3.<sup>20</sup> Rab proteins function as molecular switches that regulate the formation, transport, tethering and fusion of a variety of membrane structures by cycling between inactive GDP-bound and active GTP-bound states.<sup>21–23</sup> GTP binding to Rab proteins is mediated by a guanine exchange factor (GEF), and subsequently Rab3GAP was shown to also function as a GEF for another Rab protein, Rab18.<sup>24</sup>

Homozygous and compound heterozygous mutations in human *RAB3GAP1*, *RAB3GAP2*, and *RAB18* cause a severe developmental disorder known as Warburg micro syndrome (WARBM1, WARBM2, and WARBM3, respectively) and the somewhat milder disease phenotype called Martsolf syndrome.<sup>25-29</sup> Other patients (WARBM4) have had mutations in TBC1D20, which encodes a protein that functions as a GAP for Rab1 and Rab2.<sup>29</sup> Children with WARBM have microcephaly with severe developmental delays and seizures, ocular abnormalities including congenital cataracts and microphthalmia, and a predominantly axonal peripheral neuropathy.<sup>28,30,31</sup> There are no reports describing histopathology in WARBM, but MRI of affected children have shown predominantly frontal polymicrogyria and cerebellar atrophy.<sup>28,32</sup> Both Rottweilers with NVSD and BRT with POANV show microphthalmia, congenital cataracts and axonal peripheral neuropathy with laryngeal paralysis,<sup>7,13,33–36</sup> The spinocerebellar ataxia reported in some Rottweilers has not been observed in BRTs though both breeds show cerebellar pathology.<sup>7,13,36</sup> Cerebral cortical dysplasias have not been reported in either breed,<sup>7,13,33-36</sup> which would explain the absence in dogs of the cognitive changes and seizures reported in children with WARBM.<sup>28</sup>

A variety of progressive neurologic diseases occur in young Rottweilers and are presumed to be hereditary (Table 1).<sup>1-6,8,9</sup> The clinical histories of the RAB3GAP1: c.743delC homozygotes in the current study most closely resemble those previously reported for dogs with NVSD. In all reports of NVSD, the onset of clinical signs was around 3 months of age.<sup>7,33,36–38</sup> The initial report described respiratory difficulties in one case but laryngeal paralysis was not documented. Ataxia and weakness were the most prominent clinical signs observed. Another report emphasized cerebellar ataxia and inspiratory stridor as the major clinical signs.<sup>36</sup> Another study described signs of larvngeal paralysis and weakness with electrodiagnostic and histopathology suggesting an axonopathy in Rottweilers with a sensorimotor neuropathy beginning around 3 months of age, but did not report any lesions in the brain or spinal cord<sup>33</sup> Later studies emphasized the laryngeal paralysis and polyneuropathy seen in NVSD.<sup>35,37,39</sup> Cataracts and microphthalmia have also been reported in some cases of NVSD or polyneuropathy in Rottweilers.<sup>33–35</sup> Compared to the NVSD cases, dogs with spinal muscular atrophy and distal myopathy had earlier ages at onset; whereas most dogs with neuroaxonal dystrophy, sensory neuropathy, and leukoencephalomyelopathy had later ages at onset. One dog that was diagnosed at necropsy with neuroaxonal dystrophy presented at 15 weeks of age with laryngeal paralysis<sup>40</sup> and might have, in fact, had NVSD. Now such cases could be genotyped for the *RAB3GAP1: c.743delC* allele to clarify whether or not they should be reclassified as cases of NVSD.

The cause of an NVSD-like disease phenotype in Boxers<sup>41</sup> is currently unknown. A recent report describes POANV in Huskies with a SINE insert in *RAB3GAP1*.<sup>14</sup> Affected dogs had microphthalmia, miosis, cataracts, and persistent pupillary membranes. Voice changes and megaesophagus were reported, but as in Rottweilers, the severe laryngeal paralysis seen in BRT<sup>7</sup> was not observed. The affected Huskies survived longer than the BRT and developed a severe sensory ataxia. Histopathology revealed neuronal vacuolation and axonal neuropathy.<sup>14</sup> The occurrence of similar disease phenotypes associated with 2 different *RAB3GAP1* mutations in 3 different breeds supports the existence of a causal relationship between the mutations and the disease.

The first cases of NVSD were seen soon after the recognition of variant Creutzfeldt-Jakob disease (CJD) in humans following the bovine spongiform encephalopathy (BSE) epidemic. Variant CJD was a novel form of CJD and transmission of the BSE prion to humans was suspected.<sup>42</sup> The recognition of a spongiform encephalopathy in dogs raised concerns that it could also be a prion disease. Further studies at the time, however, did not demonstrate protease-resistant prion protein on immunohistochemistry or Western immunoblot assay<sup>7,36,37</sup> and no mutations were identified in PRNP, the gene coding for prion protein, in an affected dog (Johnson GS and O'Brien, unpublished observation). The cause of the vacuolar change in prion diseases and its relationship with the pathogenesis of the disease are not known.<sup>43</sup> Studies of sporadic CJD have shown decreased expression of a Rab recycling protein and increased activated Rab3a in Purkinje cells which may contribute to the pathogenesis of the disease.<sup>44,45</sup> Further investigation of membrane trafficking in dogs with

Disease	Age of Onset	Major Signs	Pathology
Spinal muscular atrophy <sup>4,5</sup>	4 weeks	Progressive paralysis and hypotonia	Motor neuron degeneration
Distal myopathy <sup>8</sup>	3-8 weeks	Planitgrade/palmigrade stance and weakness	Myofiber atrophy and endomysial fibrosis
Myotubular myopathy9	7-13 weeks	Weakness and hypotonia in males (x-linked)	Small muscle fibers with central nuclei and necklace fibers
Neuronal vacuolation and spinocerebellar degeneration <sup>7,36</sup>	12 weeks	Ataxia, weakness and laryngeal paralysis	Neuronal vacuolation
Neuroaxonal dystrophy <sup>1,3</sup>	1.5-4 years	Progressive sensory ataxia and nystagmus	Spheroids in sensory tracks
Distal sensorimotor neuropathy <sup>6</sup>	1.5-4 years	Progressive paralysis, hypotonia, muscle atrophy and slow NCV	Loss and thinning of myelin, axonal necrosis and neurogenic muscle atrophy
Leukoencephalomyelopathy <sup>2</sup>	3-4 years	Progressive cerebellar ataxia and weakness	Demyelination of spinal cord, brainstem, and cerebellum

 Table 1. Comparison of some presumed hereditary neurologic diseases reported in Rottweilers

*RAB3GAP1:c.743delC* could shed light on the pathogenesis of transmissible spongiform encephalopathies.

In conclusion, our data support a causal relationship between homozygosity for the RAB3GAP1:c.743delC allele and NVSD in Rottweilers and POANV in BRT as well as the RAB3GAP1 SINE insertion and POANV in Huskies.<sup>13,14</sup> We would recommend using POANV to describe all 3 diseases since it encompasses more of the signs associated with the mutations. The availability of DNA tests for the deletion allele should aid in the diagnosis of the disease and clarify the relationship between the various young onset neurologic diseases in Rottweilers. It should also permit breeders to identify carriers of the mutant allele. They can then use wise breeding strategies to avoid producing affected dogs while maintaining desirable traits and genetic diversity in their line. Finally, affected dogs could serve as a model for investigating how altered membrane trafficking leads to neurodegenerative disease.

## Footnote

<sup>a</sup> Whatman FTA<sup>®</sup> Elute cards, GE Healthcare Life Sciences, Marlborough, MA

## Acknowledgments

The authors thank all the veterinarians, breeders, and dog owners who assisted in collecting DNA samples and the clinical data.

*Conflict of Interest Declaration:* Authors declare no conflict of interest.

*Off-label Antimicrobial Declaration:* Authors declare no off-label use of antimicrobials.

#### References

1. Cork LC, Troncoso JC, Price DL, et al. Canine neuroaxonal dystrophy. J Neuropathol Exp Neurol 1983;42:286–296.

2. Gamble DA, Chrisman CL. A leukoencephalomyelopathy of rottweiler dogs. Vet Pathol 1984;21:274–280.

3. Chrisman CL, Cork LC, Gamble DA. Neuroaxonal dystrophy of Rottweiler dogs. J Am Vet Med Assoc 1984;184:464–467.

4. Shell LG, Jortner BS, Leib MS. Spinal muscular atrophy in two Rottweiler littermates. J Am Vet Med Assoc 1987;190:878–880.

5. Shell LG, Jortner BS, Leib MS. Familial motor neuron disease in Rottweiler dogs: Neuropathologic studies. Vet Pathol 1987;24:135–139.

6. Braund KG, Toivio-Kinnucan M, Vallat JM, et al. Distal sensorimotor polyneuropathy in mature Rottweiler dogs. Vet Pathol 1994;31:316–326.

7. Kortz GD, Meier WA, Higgins RJ, et al. Neuronal vacuolation and spinocerebellar degeneration in young Rottweiler dogs [see comments]. Vet Pathol 1997;34:296–302.

8. Hanson SM, Smith MO, Walker TL, et al. Juvenile-onset distal myopathy in Rottweiler dogs. J Vet Intern Med 1998;12:103–108.

9. Shelton GD, Rider BE, Child G, et al. X-linked myotubular myopathy in Rottweiler dogs is caused by a missense mutation in Exon 11 of the MTM1 gene. Skelet Muscle 2015;5:1.

10. Chrisman CL. Neurological diseases of Rottweilers: Neuroaxonal dystrophy and leukoencephalomalacia. J Small Anim Pract 1992;33:500–504.

11. Davies DR, Irwin PJ. Degenerative neurological and neuromuscular disease in young rottweilers. J Small Anim Pract 2003;44:388–394.

12. O'Brien DP, Leeb T. DNA testing in neurologic diseases. J Vet Intern Med 2014;28:1186–1198.

13. Mhlanga-Mutangadura T, Johnson GS, Schnabel RD, et al. A mutation in the Warburg syndrome gene, RAB3GAP1, causes a similar syndrome with polyneuropathy and neuronal vacuolation in Black Russian Terrier dogs. Neurobiol Dis 2016; 86:75–85.

14. Wiedmer M, Oevermann A, Borer-Germann SE, et al. A RAB3GAP1 SINE insertion in Huskies with polyneuropathy, ocular abnormalities and neuronal vacuolation (POANV) resembling human Warburg Micro Syndrome 1 (WARBM1). G3 (Bethesda) 2015;6(2):255–262.

15. Dubowitz V, Sewry CA. HIstological and histochemical stains and reactions. In: Dubowitz V, Sewry CA, Oldfors A, eds. Muscle Biopsy: A practical approach, 4th ed. St. Louis, MO: Saunders Elsevier; 2013:16–27.

16. Mizisin AP, Nelson RW, Sturges BK, et al. Comparable myelinated nerve pathology in feline and human diabetes mellitus. Acta Neuropathol 2007;113:431–442.

17. Zeng R, Coates JR, Johnson GC, et al. Breed distribution of SOD1 alleles previously associated with canine degenerative myelopathy. J Vet Intern Med 2014;28:515–521.

18. Katz ML, Khan S, Awano T, et al. A mutation in the CLN8 gene in English Setter dogs with neuronal ceroid-lipofuscinosis. Biochem Biophys Res Commun 2005;327:541–547.

19. Livak KJ. Allelic discrimination using fluorogenic probes and the 5' nuclease assay. Genet Anal 1999;14:143–149.

20. Fukui K, Sasaki T, Imazumi K, et al. Isolation and characterization of a GTPase activating protein specific for the Rab3 subfamily of small G proteins. J Biol Chem 1997;272:4655–4658.

21. Hutagalung AH, Novick PJ. Role of Rab GTPases in membrane traffic and cell physiology. Physiol Rev 2011;91:119–149.

22. Zirin J, Nieuwenhuis J, Samsonova A, et al. Regulators of autophagosome formation in Drosophila muscles. PLoS Genet 2015;11:e1005006.

23. Bhuin T, Roy JK. Rab proteins: The key regulators of intracellular vesicle transport. Exp Cell Res 2014;328:1–19.

24. Gerondopoulos A, Bastos RN, Yoshimura S, et al. Rab18 and a Rab18 GEF complex are required for normal ER structure. J Cell Biol 2014;205:707–720.

25. Aligianis IA, Johnson CA, Gissen P, et al. Mutations of the catalytic subunit of RAB3GAP cause Warburg Micro syndrome. Nat Genet 2005;37:221–223.

26. Aligianis IA, Morgan NV, Mione M, et al. Mutation in Rab3 GTPase-activating protein (RAB3GAP) noncatalytic subunit in a kindred with Martsolf syndrome. Am J Hum Genet 2006;78:702–707.

27. Bem D, Yoshimura S, Nunes-Bastos R, et al. Loss-of-function mutations in RAB18 cause Warburg micro syndrome. Am J Hum Genet 2011;88:499–507.

28. Handley MT, Morris-Rosendahl DJ, Brown S, et al. Mutation spectrum in RAB3GAP1, RAB3GAP2, and RAB18 and genotype-phenotype correlations in warburg micro syndrome and Martsolf syndrome. Hum Mutat 2013;34:686–696.

29. Liegel RP, Handley MT, Ronchetti A, et al. Loss-of-function mutations in TBC1D20 cause cataracts and male infertility in blind sterile mice and Warburg micro syndrome in humans. Am J Hum Genet 2013;93:1001–1014.

30. Graham JM Jr, Hennekam R, Dobyns WB, et al. MICRO syndrome: An entity distinct from COFS syndrome. Am J Med Genet A 2004;128a:235–245.

31. Nassogne MC, Henrot B, Saint-Martin C, et al. Polymicrogyria and motor neuropathy in Micro syndrome. Neuropediatrics 2000;31:218–221.

32. Morris-Rosendahl DJ, Segel R, Born AP, et al. New RAB3GAP1 mutations in patients with Warburg Micro Syndrome from different ethnic backgrounds and a possible founder effect in the Danish. Eur J Hum Genet 2010;18:1100–1106.

33. Mahony OM, Knowles KE, Braund KG, et al. Laryngeal paralysis-polyneuropathy complex in young Rottweilers. J Vet Intern Med 1998;12:330–337.

34. O'Brien JA. Laryngeal paralysis in dogs. In: Kirk RW, ed. Current Veterinary Therapy. Philadelphia: W.B. Saunders; 1986:789–792.

35. Salvadori C, Tartarelli CL, Baroni M, et al. Peripheral nerve pathology in two rottweilers with neuronal vacuolation and spinocerebellar degeneration. Vet Pathol 2005;42:852–855.

36. van den Ingh TS, Mandigers PJ, van Nes JJ. A neuronal vacuolar disorder in young rottweiler dogs [see comments.]. Vet Rec 1998;142:245–247.

37. Eger CE, Huxtable CR, Chester ZC, et al. Progressive tetraparesis and laryngeal paralysis in a young rottweiler with neuronal vacuolation and axonal degeneration: An Australian case. Aust Vet J 1998;76:733–737.

38. Pumarola M, Fondevila D, Borras D, et al. Neuronal vacuolation in young Rottweiler dogs. Acta Neuropathol 1999;97:192–195.

39. de Lahunta A, Summers BA. The laryngeal lesion in young dogs with neuronal vacuolation and spinocerebellar degeneration. Vet Pathol 1998;35:316–317.

40. Bennett PF, Clarke RE. Laryngeal paralysis in a rottweiler with neuroaxonal dystrophy. Aust Vet J 1997;75:784–786.

41. Geiger DA, Miller AD, Cutter-Schatzberg K, et al. Encephalomyelopathy and polyneuropathy associated with neuronal vacuolation in two Boxer littermates. Vet Pathol 2009;46:1160–1165.

42. Will RG, Ironside JW, Zeidler M, et al. A new variant of Creutzfeldt-Jakob disease in the UK. Lancet 1996;347:921–925.

43. Gunn TM, Carlson GA. RML prions act through Mahogunin and Attractin-independent pathways. Prion 2013;7:267–271.

44. Gawinecka J, Cardone F, Asif AR, et al. Sporadic Creutzfeldt-Jakob disease subtype-specific alterations of the brain proteome: Impact on Rab3a recycling. Proteomics 2012;12:3610–3620.

45. Ferrer I, Puig B, Blanco R, et al. Prion protein deposition and abnormal synaptic protein expression in the cerebellum in Creutzfeldt-Jakob disease. Neuroscience 2000;97:715–726.